Outbreak of *Mycobacterium chelonae* Infection Associated with Tattoo Ink


ABSTRACT

BACKGROUND

In January 2012, on the basis of an initial report from a dermatologist, we began to investigate an outbreak of tattoo-associated *Mycobacterium chelonae* skin and soft-tissue infections in Rochester, New York. The main goals were to identify the extent, cause, and form of transmission of the outbreak and to prevent further cases of infection.

METHODS

We analyzed data from structured interviews with the patients, histopathological testing of skin-biopsy specimens, acid-fast bacilli smears, and microbial cultures and antimicrobial susceptibility testing. We also performed DNA sequencing, pulsed-field gel electrophoresis (PFGE), cultures of the ink and ingredients used in the preparation and packaging of the ink, assessment of source water and faucets at tattoo parlors, and investigation of the ink manufacturer.

RESULTS

Between October and December 2011, a persistent, raised, erythematous rash in the tattoo area developed in 19 persons (13 men and 6 women) within 3 weeks after they received a tattoo from a single artist who used premixed gray ink; the highest occurrence of tattooing and rash onset was in November (accounting for 15 and 12 patients, respectively). The average age of the patients was 35 years (range, 18 to 48). Skin-biopsy specimens, obtained from 17 patients, showed abnormalities in all 17, with *M. chelonae* isolated from 14 and confirmed by means of DNA sequencing. PFGE analysis showed indistinguishable patterns in 11 clinical isolates and one of three unopened bottles of premixed ink. Eighteen of the 19 patients were treated with appropriate antibiotics, and their condition improved.

CONCLUSIONS

The premixed ink was the common source of infection in this outbreak. These findings led to a recall by the manufacturer.

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Since 2003, a growing number of published case reports have linked tattooing with localized infections due to atypical mycobacteria.1-13 *Mycobacterium chelonae* is a rapidly growing form of nontuberculous mycobacteria; overall, it is an uncommon cause of cutaneous infections.14 We describe an investigation of an outbreak involving 19 persons with presumed *M. chelonae* infection after receiving a tattoo from a single artist who used a premixed ink that was contaminated before distribution. Previous outbreaks were associated with dilution and contamination of ink at the tattoo parlor.

**METHODS**

**INDEX PATIENT**

On January 4, 2012, we and colleagues at the Monroe County Department of Public Health began investigating an outbreak on the basis of a report from a dermatologist of the development of a persistent granulomatous rash on the arm of a person who had received a tattoo on that area in October 2011. This index patient was a previously healthy 20-year-old man who had a history of multiple tattoos without any health problems. He was treated unsuccessfully with glucocorticoids by his primary care provider before being referred to a dermatologist. Histopathological examination and culture of a biopsy specimen confirmed the presence of *M. chelonae*.

**EPIDEMIOLOGIC INVESTIGATION**

After the interview with the index patient, investigators at the Monroe County Department of Public Health learned that similar reactions had developed in other persons who had obtained tattoos from the same artist at the same tattoo parlor. During a site visit, investigators interviewed the tattoo artist and learned that since May 2011, he had been using a new, hand-blended, diluted black ink (gray wash) that contained pigment, distilled water, witch hazel, and glycerin. Gray wash is used to achieve shading and a three-dimensional quality in tattoos; it is a prominent component of portrait and photography tattoos, which are currently popular.

According to the artist, the manufacturer diluted the black ink with distilled water before packaging and shipping the ink. This premixed ink was prepackaged in three dilution strengths per set: a 30%, or light, wash (i.e., greatest dilution); a 60%, or medium, wash (i.e., intermediate dilution); and a 90%, or dark, wash (i.e., least dilution). The tattoo artist originally purchased the ink in April 2011 at a tattoo trade show in Arizona, and he later ordered an additional supply. The product labeling contained no lot number or other unique manufacturing identification. He stopped using the premixed ink in December 2011.

To identify cases of infection, a number of steps were taken. First, a regional pathology laboratory used by many dermatologists was alerted to report any cases of ink-associated granulomatous reactions. Second, a list of patrons who were tattooed between May and December 2011 was obtained from the tattoo artist. The artist did not maintain detailed records that included the type of ink that was used with each patron. Third, all local tattoo parlors within Monroe County were identified, contacted by telephone, and queried about ink products used in their establishments as well as any reports of rashlike illnesses. No other parlor used this premixed ink, and no rashlike illnesses were reported. Finally, exposed patrons were contacted by telephone and interviewed, with the use of a standard questionnaire, about receiving a tattoo and any subsequent rashlike illness.

An environmental health assessment suggested that best practices were generally observed at the parlor where the index patient received his tattoo, and no concerns about hygiene were noted. In particular, the artists used sterile instruments, wore clean disposable gloves, poured ink into single-use containers, and provided appropriate aftercare to the tattooed site. Importantly, no dilution or mixing of inks at the parlor occurred, and the artist avoided contamination of ink from tap water at the facility. These practices were confirmed by all interviewed patrons.

The Food and Drug Administration (FDA) inspected the Arizona-based manufacturer and distributor of the ink. The FDA collected samples of ink and of the ingredients used in its preparation and packaging, and the Centers for Disease Control and Prevention (CDC) cultured them.

The clinical findings in all but two patients were evaluated at the University of Rochester Medical Center; of the two patients with findings that were not evaluated, one had follow-up in another state and another declined evaluation and treatment. Patients were evaluated by dermatologists, who performed skin biopsies of the
tattooed lesions; tissue specimens were then sent for histologic and microbiologic analysis. Patients were referred to an infectious-disease expert for follow-up care and offered empirical treatment with appropriate antimicrobial agents.

**HISTOLOGIC AND MICROBIOLOGIC INVESTIGATION**

Laboratory investigations were conducted at the University of Rochester Medical Center, the Wadsworth Center of the New York State Department of Health, and the CDC. Samples from clinical isolates, one opened bottle of 60% premixed gray ink wash and three unopened bottles (one containing 30% wash, one containing 60% wash, and one containing 90% wash), and water and swabs from faucets (biofilm) were submitted for various microbiologic tests, including acid-fast bacilli smears, cultures and antimicrobial sensitivity profiles, DNA sequencing, and pulsed-field gel electrophoresis (PFGE), according to standard protocols.15-20

**RESULTS**

**EPIDEMIOLOGIC INVESTIGATION**

In this outbreak of *M. chelonae* infections associated with tattoos, 19 cases were identified: 14 confirmed cases, 4 probable cases, and 1 suspected case (see Table S1 in the Supplementary Appendix, available with the full text of this article at NEJM.org). On the basis of a list of 167 patrons provided by the tattoo artist, the attack rate increased from zero during the period from May through August to a peak of 65% in November (Fig. 1). Rashes were confined to areas of the skin tattooed with the premixed gray ink (Fig. 2). The average age of the 13 men and 6 women with *M. chelonae* infection was 35 years (range, 18 to 48).

**HISTOLOGIC AND MICROBIOLOGIC INVESTIGATION**

Biopsy specimens were obtained from 17 of the 19 patients: sparse lymphohistiocytic infiltrates in the upper dermis were observed in 12 specimens, granulomas in 5, and acid-fast organisms in 2 (Fig. S1 in the Supplementary Appendix). *M. chelonae* was isolated in 14 of the 17 tissue specimens and confirmed by DNA sequencing.

Antimicrobial susceptibility studies for atypical mycobacteria are not routinely performed in the United States; however, these studies were performed in two patients. In the first patient, *M. chelonae* was sensitive to clarithromycin, doxycycline, and linezolid; had intermediate sensitivity to ciprofloxacin; and was resistant to cefoxitin. In the second patient, *M. chelonae* was sensitive to clarithromycin and doxycycline, had intermediate sensitivity to linezolid, and was resistant to ciprofloxacin and cefoxitin.
The CDC results confirmed the isolation of M. chelonae and showed an indistinguishable PFGE pattern in 11 clinical isolates and one of three unopened bottles of ink (i.e., the 30% gray wash) (Fig. S2 in the Supplementary Appendix). M. chelonae was not isolated from the water or faucet in the tattoo parlor.

**Infection-Control Measures**

We collected the remaining bottles of premixed ink from the tattoo parlor. We also provided information and education to both tattoo artists and patrons at a large tattoo trade show hosted in Rochester, New York. The materials provided included information on New York State law, which bans tattooing of persons younger than 18 years of age, regardless of whether parental consent is given; tattooing best practices, including the use of sterile products and aseptic techniques as well as appropriate aftercare; risks associated with tattooing; signs of a reaction from a tattoo; and steps to be taken if a reaction to a tattoo occurs.

The CDC issued a nationwide alert about the outbreak. The manufacturer voluntarily recalled the ink. Appropriate antimicrobial therapy (macrolides at first, based on the recommendations from relevant prior case reports, and then macrolides, doxycycline, or both, once sensitivities were available) was initiated in 18 of the 19 patients, and their condition improved, although the speed of recovery in each patient varied.

**Discussion**

In this outbreak in Rochester, New York, cutaneous M. chelonae infection developed in 19 patrons of a tattoo parlor, with culture confirmation in 14, after tattooing performed with the use of a premixed gray ink manufactured in Arizona. The epidemiologic, histologic, and microbiologic evidence implicated the ink, not the practices of the tattoo parlor, as the common source of infection.

Previous studies have linked M. chelonae to other procedures. however, only in the past decade have cutaneous infections with nontuberculous mycobacteria been associated with tattooing, with Wolf and Wolf describing the first case in 2003. According to their report, cultures were negative, but polymerase-chain-reaction analysis confirmed the diagnosis; they were unable to confirm the mycobacterial species. Goldman et al. reported on a case series involving 48 patients with M. chelonae infections linked to tattooing in France. In that outbreak, M. chelonae was isolated in 13 patients; no PFGE analysis was performed. M. chelonae was isolated from opened, but not unopened, bottles of ink. In the United States, Drage et al. described an outbreak of M. chelonae infections involving 6 patients with tattoos. In that case series, cultures confirmed M. chelonae infection in 3 patients.

Previous studies, although not testing specifically for mycobacteria, have shown that unopened stock bottles of tattoo ink may contain bacteria that are pathogenic in humans despite claims of sterility. In the current study, mycobacteria were detected in both opened and unopened bottles of ink. These findings suggest that mycobacteria may be another potential contaminant, especially if the ink is diluted with nonsterile water before distribution.

A major limitation of this investigation was the lack of detailed records on the patrons of the tattoo parlor, which affected the assessment of exposure. The tattoo artist was uncertain about which patron received which batch of ink and about the dilution of the premixed gray ink used. It seems reasonable to suppose that the first batch was used during the earlier exposure period, since no cases of infection developed until October; this suggests that only the second batch was con-
taminated. The likelihood of overlooked cases was minimized through the active surveillance efforts undertaken by the public health department.

Our findings have at least two implications. First, since tattooing has become more popular over time, the incidence of cutaneous mycobacterial infections may be underestimated, given the lack of both routine testing and mandated reporting. Second, although contamination in tattoo parlors has been implicated in previous outbreaks, our investigation of this outbreak shows that premixed ink contaminated before distribution poses a risk to public health, which may suggest the need for enhanced oversight of not just tattooing but also the inks used in tattooing to ensure public safety.

The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the Centers for Disease Control and Prevention or the Agency for Toxic Substances and Disease Registry. Disclosure forms provided by the authors are available with the full text of this article at NEJM.org.

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**REFERENCES**